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A BICENTENNIAL OF BETULIN

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Key Word Index—Betulin; chemotaxonomy of angiosperms; history; identification; isolation; pharmacology; physical properties; reactivity; review; utilisation.

Abstract—The literature on the history, identification, determination, isolation and distribution in plants of betulin as well as its technological applications and its pharmacology is reviewed and the chemotaxonomy of the angiosperms in relation to betulin is discussed.

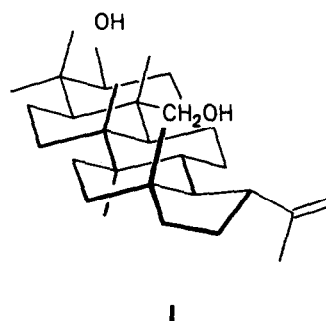
History and Introduction

With the oldest reference on betulin (1) dating back to 1788 [1] this substance certainly constitutes one of the first natural products isolated from plants and detailed investigations including elemental analysis were described in 1876 by Hausmann [2]. Betulin, as a pentacyclic triterpene with the lupane skeleton, is also named betulinol, trochton, birkenkampher and (coryli-) resinol. It is referenced in 'Chemical Abstracts' as lup-20(29)-en-3,28-diol, (3 β). Systematic names are 5a.5b.8.8.11a-pentamethyl-3a-hydroxymethyl-1-isopropenyl-eicosahydro-1H-cyclopenta[a]-chrysen-9-ol or (3a*S*)-9c-hydroxy-5a.5b.8.8.11a-pentamethyl-3a-hydroxymethyl-1*t*-iso-propenyl-(7a*H*.11*bH*.13a*H*.13*bH*)-eicosahydro-1H-cyclopenta[a]chrysen [3]. The latest review on betulin with focus on reactions and uses appeared in 1981 [4].

Isolation and detection

Betulin is isolated primarily from barks [5] either via solvent extraction or sublimation [6] and purified by column chromatography or recrystallization; procedures for the isolation of betulin from birch bark in appreciable amounts [2, 7–15] are well described. Procedures for the isolation from peat [16, 17] lignite [18] and archeological specimens [13, 19] or wood pulp extracts [20, 21] are found as well. For the extractive isolation of betulin on an industrial scale from birch bark waste, various solvents have been compared [21–25]; the azeotrope of 2-butanol–water was found to be the best [22]. Some authors suggest the purification via its diacetate [6, 24] followed by hydrolysis. From our experience methods relying only on recrystallization for purification of betulin are insufficient. Betulin is also used in geochemistry to trace variations in provenance, fixation and diagenesis of organic matter deposited in sediments along the continental margin [26].

For the detection of betulin combinations of the procedures mentioned below have been utilized:



TLC [19, 27–29]. Combined with the use of spray reagents and densitometers [25] were described as well as R_f -structure relationships [30] and TLC using AgNO₃-silica gel [29, 31]. From our experience detections of betulin based only on TLC should be critically re-evaluated: far too many triterpenoids exhibit similar R_f -values and behaviour towards spray reagents [30].

HPLC. The use of normal phase as well as C₁₈-reversed phase columns are described in detail [32]. Although betulin can be detected by UV at 205–215 nm and with refractive index detectors, the sensitivities of both methods are low.

GC. The techniques for the separation of betulin, its diacetate and TMSi-ethers have been investigated on packed [29, 33–35] and capillary columns [28, 36].

Droplet Counter Current Chromatography (DCCC). The separation of betulin from betulinic acid, β -amyrin and cholesterol was achieved by this method [37].

Physical properties

Melting points of betulin are reported over a range of 250–261° with the higher values obtained after sublimation of the recrystallized product [2, 38, 39]. The lower values can be explained by the crystallization with 1 mol of ethanol [9, 14]. Some authors further indicate mixed melting points, e.g. of eutectics with saccharin [27].

Table 1. Distribution of betulin in angiosperms

Plant name	Part used	Analysis	Quantity
<i>Acacia modesta</i> [118]	b	IR, mp, TLC	
<i>Acacia nilotica</i> [119]	rb	IR, mp, TLC	
<i>Acacia senegal</i> [120]	r	IR, mp, TLC	
<i>Acacia tortilis</i> [121]	b	IR, mp, TLC	
<i>Acinos thymoides</i> [122]			0.03
<i>Acrotrema arnoethianum</i> [183]	l	IR, mp, TLC	0.065
<i>Akebia quinata</i> [123]	st		
<i>Alangium lamarckii</i> [124]	s		
<i>Alnus barata</i> [125]	b	mp, ORD	
<i>Alnus crispa</i> [83]	b	GC	<5
<i>Alnus glutinosa</i> [126]	b	mp	
<i>Alnus maximowiczii</i> [127, 128]	b, r		
<i>Alnus nepalensis</i> [129]	b	mp, MS, ORD	0.23
<i>Alnus rubra</i> [130]	b, w	mp, ORD	0.036
<i>Alnus rugosa</i> [83]	b, f, l	GC	<5
<i>Alnus subcordata</i> [131]	b	mp	
<i>Alphitonia whitei</i> [132]	w	mp	0.0012
<i>Alstonia scholaris</i> [133]	l	IR, mp, TLC	
<i>Amoora canarana</i> [134]			
<i>Amanoa oblongifolia</i> [93]	b	IR, mp, MS, ORD	0.00004
<i>Anisomeles ovata</i> [135]	ae	IR, MS, NMR, TLC	0.001
<i>Aspidixia articulata</i> [136]			
<i>Asteracantha longifolia</i> [137]	r	IR, mp, MS, NMR, UV	
<i>Avicennia alba</i> [138]	b, l	IR, MS, mp, ORD	0.01, 0.012
<i>Avicennia officinalis</i> [138]	b, l	IR, MS, mp, ORD	0.014–0.027, 0.006–0.01
<i>Befaria glutinosa</i> [139]	fl		
<i>Betula</i> [1, 2, 5, 9, 140, 141, 142]	b, ob	mp, IR, mp, ORD	10.0
<i>Betula alba</i> [11, 12, 109, 143]	b, ob	mp	14.0–20.0
<i>Betula alleghaniensis</i> [148]	b	mp	
<i>Betula bojpattra</i> [153]	b	mp	10–20
<i>Betula carpatica</i> [153]	b	mp	10–20
<i>Betula cordifolia</i> [82]	ob	GC/MS	6.8
<i>Betula costata</i> [144, 153]	b	IR, mp	<5
<i>Betula dahurica</i> [145]	b+br		
<i>Betula daurica</i> [127]	b		
<i>Betula ermanii</i> [144]	b	IR, mp	
<i>Betula lepta</i> [148, 153]	b	mp	0.05, <5
<i>Betula luminifera</i> [153]	b	mp	<5
<i>Betula hūea</i> [153]	b	mp	5–10
<i>Betula mandschurica</i> [146]	ob		
<i>Betula maximowicziana</i> [147]	ob	IR, mp, MS	
<i>Betula papyrifera</i> [82, 83, 148, 153]	b, ob	mp, GC, GC/MS	10–20, <30
<i>Betula pendula</i> L. [14, 39]	b, ob	mp	23, 36
<i>Betula pendula</i> [27]	b	IR, mp, TLC	1.08
<i>Betula platyphylla</i> [28, 149]	b, ob	MS, NMR	
<i>Betula populifolia</i> [82, 153]	b, ob	mp, GC/MS	10–20
<i>Betula pubescens</i> [153]	b	mp	10–20
<i>Betula pubescens</i> [27]	b	IR, mp, TLC	0.36
<i>Betula sanbergii</i> [153]	b	mp	5–10
<i>Betula schmidtii</i> [153]	b	mp	≤10
<i>Betula utilis</i> [78, 107, 150, 151]	b, ob	IR, mp, ORD, TLC	23.5
<i>Betula utilis</i> [153]	b	mp	5–10
<i>Betula verrucosa</i> [22, 25, 152, 153]	b, ob	mp, TLC	10–20
<i>Betula verrucosa</i> [154]	ob	GC/MS	34.0
<i>Betula x caerulea</i> [82]	ob	GC/MS	11.4
<i>Buxus microphylla</i> var <i>intermedia</i> [155]	b+l	IR, mp	
<i>Buxus microphylla</i> var <i>suffructuosa</i> [156]	wp	mp	
<i>Buxus microphylla</i> [57]	l+st		
<i>Buxus papillosa</i> [158]	l	IR, mp, MS, NMR	
<i>Buxus sempervirens</i> [159]	l		
<i>Callistemon lanceolatus</i> [160]	l	mp, ORD	
<i>Canavalia bonariensis</i> [161]	r	IR, mp, NMR, TLC	

Table 1. (Continued)

Plant name	Part used	Analysis	Quantity
<i>Capparis sepiaria</i> [162]	l		
<i>Caraipa densifolia</i> [163]	b	IR, mp, TLC	0.003
<i>Carlina corymbosa</i> var <i>globosa</i> [164]	ae	GC, IR, mp, MS, NMR	
<i>Carpinus betulus</i> [38, 39, 165]	b	mp, ORD	0.04
<i>Cassia holosericeae</i> [166]	ae		
<i>Cassia siamea</i> [167]	fl	IR, mp, MS, NMR, ORD	0.0005
<i>Cassia siamea</i> [168]	b	IR, mp, TLC	
<i>Castanea sativa</i> [169, 170]	l	mp, ORD	0.4
<i>Castanopsis concinna</i> [171]	st	IR, mp	0.00067
<i>Castanopsis eyrei</i> [171]	st	mp	0.0009
<i>Ceanothus americanus</i> [134]	rb		
<i>Celtis australis</i> [151]	b	IR, mp, NMR, TLC	0.12
<i>Cistus laurifolius</i> [172]	wp	IR, MS, mp	
<i>Clerodendron inerme</i> [173]	st		
<i>Clinacanthus nutans</i> [174]		IR, MS, NMR	
<i>Clinopodium vulgare</i> [175, 176]	ae, l		
<i>Comptonia peregrina</i> [83]	l, st, r	GC	< 5
<i>Cordia obliqua</i> [177]	s		
<i>Cornus capitata</i> [178]	st	IR, MS, NMR, TLC	0.0008
<i>Corylus avellana</i> [39, 179]	b	mp	0.13
<i>Dalbergia sericea</i> [180]	b	mp, ORD	0.0033
<i>Desmodium pulchellum</i> [181]	r	IR, mp	
<i>Dicliptera roxburghiana</i> [182]	l		
<i>Dillenia andamanica</i> [183]	l	IR, mp, TLC	0.082
<i>Dillenia aurea</i> [183]	l	IR, mp, TLC	0.045
<i>Dillenia bracteata</i> [183]	l	IR, mp, TLC	0.07
<i>Dillenia indica</i> [183, 184, 185, 186]	b, fr, l	IR, mp, ORD, TLC	0.105
<i>Dillenia pentagyna</i> [183, 187]	b, l	IR, mp, MS, NMR, TLC	0.15
<i>Dillenia peregrina</i> [186]	b	IR, mp, ORD, TLC	
<i>Dillenia retusa</i> [183]	l	IR, mp, TLC	0.068
<i>Dillenia scabralla</i> [183]	l	IR, mp, TLC	0.12
<i>Diospyros abyssinica</i> [188]	b	TLC	
<i>Diospyros argentea</i> [189]	b	TLC	
<i>Diospyros bioindensis</i> [190]	b	IR, mp, TLC	
<i>Diospyros buxifolia</i> [191, 192]	hw	TLC	0.03
<i>Diospyros canaliculata</i> [188]	b	TLC	
<i>Diospyros chevalieri</i> [188]	b	TLC	
<i>Diospyros chloroxylon</i> [193]	r		
<i>Diospyros cinnabarina</i> [188, 190]	b, w	TLC	
<i>Diospyros consolatae</i> [194, 195]	l	GC/MS, mp, NMR, TLC	0.421
<i>Diospyros crassiflora</i> [188]	b	TLC	
<i>Diospyros dendo</i> [188]	b, w	TLC	≤ 0.006
<i>Diospyros discolor</i> [189, 212]	b, fr, w	IR, mp, ORD, TLC	0.05, 0.08
<i>Diospyros ebenaster</i> [196]	r		
<i>Diospyros ebenum</i> [197, 198, 199]	b, hw, l	IR, mp	≤ 0.053
<i>Diospyros elliptifolia</i> [200]	b		
<i>Diospyros fragrans</i> [188]	w	TLC	
<i>Diospyros gabunensis</i> [188]	b	TLC	
<i>Diospyros gracilescens</i> [188, 201]	b, w	TLC	≤ 0.012
<i>Diospyros guianensis</i> [202]	b		
<i>Diospyros hirsuta</i> [203]	b, w, fr	mp, TLC	0.2, 0.11, 0.008
<i>Diospyros hoylena</i> [188]	b, w	TLC	0.011, 0.019
<i>Diospyros ismaili</i> [189]	fl	IR, mp, MS, NMR, UV	0.011
<i>Diospyros iturensis</i> [188]	b, w	TLC	≤ 0.01
<i>Diospyros kaki</i> [204]	b		
<i>Diospyros kaki</i> var. <i>sylvestris</i> [205]	r + w		
<i>Diospyros kamerunensis</i> [188]	b	TLC	
<i>Diospyros kirkii</i> [206]	rb		
<i>Diospyros longiflora</i> [188]	b, w	TLC	≤ 0.009
<i>Diospyros lotus</i> [189]	b, w	TLC	
<i>Diospyros maingayi</i> [189]	b, w	IR, mp, MS, NMR, UV	0.05, 0.005
<i>Diospyros mannii</i> [207]	b	IR, mp, MS, NMR, ORD, UV	0.005

Table 1. (Continued)

Plant name	Part used	Analysis	Quantity
<i>Diospyros maritima</i> [208]	r	IR, mp	0.00026
<i>Diospyros melanoxylon</i> [209–212]	b, f, l, w	IR, mp, ORD	≤ 0.5
<i>Diospyros mespiliformis</i> [188]	b	TLC	
<i>Diospyros monobuttensis</i> [188]	b	TLC	
<i>Diospyros montana</i> [213–215]	b, p, w	IR, MS, mp, NMR, UV	0.03
<i>Diospyros moonii</i> [203]	w	mp, TLC	0.056
<i>Diospyros obliquifolia</i> [190]	b	TLC	
<i>Diospyros oblongifolia</i> [203]	fr	mp, TLC	0.1
<i>Diospyros peregrina</i> [212, 215, 216]	fr, l, p	IR, mp, ORD	1.0
<i>Diospyros peregrina</i> [217, 218]	f, l	IR, mp, ORD, TLC	
<i>Diospyros quaesita</i> [203]	b, w	mp, TLC	0.37, 0.05
<i>Diospyros rotundifolia</i> [219]	st		
<i>Diospyros Sanza-Minika</i> [192]	w	TLC	0.035
<i>Diospyros Sanza-Minika</i> [188]	b	TLC	
<i>Diospyros siamang</i> [189]	b, w	IR, mp, MS, NMR, UV	0.03, 0.01
<i>Diospyros siderophylla</i> [220]	b		
<i>Diospyros singaporensis</i> [189]	b, w	IR, mp, MS, NMR, UV	0.02, 0.013
<i>Diospyros spinescens</i> [203]	b, w	mp, TLC	0.16, 0.09
<i>Diospyros sumatrana</i> [189]	b, w	IR, mp, MS, NMR, UV	0.02
<i>Diospyros thwaitesii</i> [203]	w	mp, TLC	0.055
<i>Diospyros tomentosa</i> [191]			
<i>Diospyros variegata</i> [192]	hw	TLC	0.008
<i>Diospyros verrucosa</i> [221]	b+r		
<i>Diospyros walkeri</i> [203]	b, w	mp, TLC	0.08, 0.03
<i>Diospyros wallachi</i> [189]	b, w	IR, mp, MS, NMR, UV	0.05, 0.06
<i>Diospyros zenkeri</i> [188]	b, w	TLC	0.067, 0.007
<i>Dipteryx alata</i> [222]	b	IR, mp, NMR	
<i>Dipteryx odorata</i> [223]	b	IR, mp, MS, NMR	0.0066
<i>Duboisia leichhardtii</i> [224]	l		
<i>Ecbolium linneenum</i> [225]			
<i>Ehretia laevis</i> [226]	b	IR, mp	0.016
<i>Enicostema littorale</i> [227]	wp		
<i>Erica arborea</i> [228]	b+l		
<i>Euclea divinorum</i> [229, 231]	r	IR, mp, MS, NMR, TLC	
<i>Euclea kellau</i> [230]	br	IR, mp, NMR, ORD	
<i>Euclea natalensis</i> [231, 232]		IR, MS, NMR	
<i>Eucommia ulmoides</i> [233]	b	IR, mp, MS	
<i>Euphorbia broteri</i> [234]	ae	IR, mp, MS, NMR, ORD	
<i>Euphorbia lathyris</i> [235]	st		
<i>Euphorbia paralias</i> [236]	pl		
<i>Euphorbia paralias</i> [237]	ae	mp, TLC	0.03
<i>Euptelea polyandra</i> [238]			
<i>Fagonia cretica</i> [134]			
<i>Fagus silvatica</i> [239]	b	mp, ORD	0.0012
<i>Flindersia pimenteliana</i> [240]	b+l		
<i>Fraxinus americana</i> [83]	l, r	GC	20, 5–10
<i>Givotia rottleriformis</i> [241]	b	IR, mp, MS, NMR, TLC, ORD	0.35
<i>Glechoma hirsuta</i> [122]			0.01
<i>Gmelina arborea</i> [242]			
<i>Grewia asiatica</i> [243, 244]	b	IR, NMR, ORD, TLC	0.15
<i>Grewia bicolor</i> [245]	wp		
<i>Grewia flavescens</i> [246]	r		
<i>Grewia tenax</i> [247]	b	IR, mp, TLC	
<i>Guazuma tomentosa</i> [248]	b	mp, MS, NMR, ORD	0.0017
<i>Gymnosporia montana</i> [249]	l	mp, MS, TLC	
<i>Gymnostachyum latifolium</i> [250]	st	IR, mp, TLC	
<i>Haplophyton cimicidum</i> [251]	pl		
<i>Hedyotis lawsoniae</i> [252]	l+t	GC, GC/MS, NMR	0.00012
<i>Helichysum dendroidenum</i> [253]	l	mp	
<i>Holacantha emoryi</i> [254]			
<i>Ilex verticillata</i> [83]	b	GC	20

Table 1. (Continued)

Plant name	Part used	Analysis	Quantity
<i>Iris pallasi</i> var. <i>chinensis</i> [255]	s	IR, mp, MS, NMR	
<i>Ixora chinensis</i> [256]	l		0.026
<i>Jasminum flexile</i> [257]	l		
<i>Jasminum humile</i> [257]	l		
<i>Jasminum multiflorum</i> [257]	l		
<i>Jasminum sambac</i> [257]	l		
<i>Juglans regia</i> [258]	b		
<i>Klenia articulata</i> [259]	ap	IR, MS	0.005
<i>Lasianthus chinensis</i> [260]	l		
<i>Lawsonia alba</i> [261]	b	MS, mp, ORD	
<i>Leptospermum scoparium</i> [262]	b	mp	
<i>Lithocarpus cornea</i> [43]	st	IR, mp, MS, NMR	0.00016
<i>Lithocarpus elizabethae</i> [263]	l+st		
<i>Lithocarpus haipinii</i> [263]	l+st		
<i>Lithocarpus polystachya</i> [264]	st	mp	0.000044
<i>Lophopetalum rigidum</i> [265]	b		0.013
<i>Lophopetalum toxicum</i> [134]	b		0.7-0.8
<i>Lumitzera racemosa</i> [266]	b, l	GC, mp, TLC	0.011, 0.25
<i>Macaranga peltata</i> [267]	hw	IR, mp, TLC, UV	0.002
<i>Maytenus octogona</i> [268]	ae		
<i>Melaleuca leucadendron</i> [269]	l, st	IR, mp, TLC	0.0014, 0.0008
<i>Melodinus australis</i> [270]	ae		
<i>Melyanthes trifoliata</i> [72, 134]			0.006
<i>Micromeria benthami</i> [271]		mp	
<i>Mitrephora heyneana</i> [272]	b		
<i>Myrica pensylvanica</i> [83]	st, l	GC	10
<i>Nepeta</i> [117]	wp		0.033
<i>Nepeta aragonensis</i> [273]	ae	GC, IR, mp, NMR, ORD	
<i>Nerium oleander</i> [41]	l	NMR, TLC	0.0004
<i>Neuracanthus sphaerostachys</i> [274]		IR, mp, TLC	
<i>Nuytsia floribunda</i> [134]	ae		
<i>Origanum compactum</i> [275]	ae	IR, mp, MS, NMR	0.009
<i>Orthosphenia mexicana</i> [276]	ae	IR, MS, NMR	
<i>Ougeinia dalbergioides</i> [277, 278]	b	mp, ORD	
<i>Parthenium hysterophorus</i> [279]	l		
<i>Phillyrea latifolia</i> [228]	b+l		
<i>Phlogacanthus thyrsoiflorus</i> [116]	l	IR, MS, mp, ORD, TLC	
<i>Phyllanthus reticulatus</i> [280]	r	IR, mp, MS, NMR, UV	
<i>Platanus hybrida</i> [281]	b	mp	
<i>Polygonum perforliatum</i> [282]			
<i>Porteresia coarctata</i> [283]	lc	GC	≤ 11.0
<i>Premna integrefolia</i> [284]	b	IR, mp, NMR	
<i>Psorospermum tenuifolium</i> [285]	rb	IR, mp, MS, NMR, TLC	0.01
<i>Psychotria adenophylla</i> [286]	l		
<i>Pterocarpus santalinus</i> [287]	l	IR, mp, TLC	
<i>Pyrus communis</i> [11]	t		
<i>Quercus bambusaefolia</i> [288]	st	IR, mp	0.00015
<i>Quercus championi</i> [288]	st	IR, mp	0.00067
<i>Quercus myrsinaefolia</i> [288]	st	IR, mp	0.00006
<i>Quercus suber</i> [45]	c	IR, MS, NMR, UV	0.0011
<i>Quercus suber</i> [289-292]	b	mp	0.6-0.7
<i>Rheedia gardneriana</i> [293]			
<i>Rhizophora mucronata</i> [294]	l	GC	
<i>Rhododendron adamsii</i> [295]	sh		
<i>Rhododendron anthopogon</i> [296]	st		
<i>Rhododendron ledebourii</i> [297]	l+st		
<i>Rhododendron luteum</i> [295]	sh		
<i>Rhododendron schlippenbachii</i> [295]	sh		
<i>Rhodomyrtus tomentosa</i> [298]	l, st	IR, mp, MS	0.002, 0.001
<i>Rosmarinus officinalis</i> [299]		NMR	
<i>Roylea elegans</i> [300]	l		
<i>Rzedowskia tolangtuensis</i> [301]	ae		

Table 1. (Continued)

Plant name	Part used	Analysis	Quantity
<i>Saccharum officinarum</i> [302]			
<i>Salvia beckeri</i> [258]	ap		
<i>Salvia mexicana</i> [303]	wp	IR, MS, NMR	
<i>Salvia officinalis</i> [304]	l	GC	0.0015
<i>Sambucus nigra</i> [134, 305]	b	mp	0.011
<i>Sarcococca pruniformis</i> [306, 307]	l, wp	IR, mp, MS, ORD	0.05
<i>Sarracenia flava</i> [114]	r		
<i>Schleichera oleosa</i> [286]	b		
<i>Scirpus fluviatilis</i> [308]	rh	IR, mp, MS, ORD	0.43
<i>Scirpus tuberosus</i> [309]	r	IR, mp, NMR	
<i>Solanum aethiopicum</i> [310]	be	IR, mp	0.0025
<i>Solanum laciniatum</i> [311]	st		
<i>Solanum wrightii</i> [311]	st		
<i>Sorbus aucuparia</i> [134]	b		
<i>Sorbus americana</i> [83]	b, l	GC	10
<i>Stenocereus thurberi</i> [29]	wp	GC, mp, ORD, TLC	
<i>Sterculia foetida</i> [312]	b	mp, ORD	0.005
<i>Sterculia urens</i> [313]	b	IR, mp	0.005
<i>Strobilanthes cusia</i> [314]	r		
<i>Tectona grandis</i> [315]	b	IR, mp, MS, NMR	
<i>Tetracera akara</i> [183]	l	IR, mp, TLC	0.045
<i>Tetracera indica</i> [183]	l	IR, mp, TLC	0.01
<i>Tetracera sarmentosa</i> [183]	l	IR, mp, TLC	0.07
<i>Tetracera scandens</i> [183]	l	IR, mp, TLC	0.03
<i>Thevetia peruviana</i> [316]			
<i>Thymelea hirsuta</i> [317]			
<i>Tricholepis glaberrima</i> [318]	wp	IR, MS, NMR, TLC	
<i>Trochodendron</i> [134]	b, l, w		
<i>Trochodendron aralioides</i> [319]	l		
<i>Viscum album</i> [320]	l+st		
<i>Viscum articulatum</i> [321]	wp	GC, mp, MS, TLC	
<i>Wisteria floribunda</i> [322]	b		
<i>Woodfordia fruticosa</i> [323]	l	IR, mp, ORD	0.004
<i>Zataris multiflora</i> [324]	l+st+fl		
<i>Zizyphus jujuba</i> [134]	s		
<i>Zizyphus mauritiana</i> [325]	st		
<i>Zizyphus sativa</i> [326]	b		
<i>Zizyphus spinosus</i> [327]	s		

ae: Aerial parts; ar: arterial parts; b: bark; be: berries; br: branches; c: cork; fl: flowers; fr: fruit; hw: heartwood; l: leaves; lc: lipid constituents; ob: outer bark; p: pulp; r: roots; rb: root bark; rh: rhizoma; s: seeds; sh: shoots; st: stems; t: twigs; w: wood; wp: whole plant.

NMR. ^1H NMR spectra at betulin have been reported [40, 41] and the chemical shifts for the methyl groups have been assigned [42, 43]. ^{13}C NMR spectra were fully interpreted [41, 44, 45].

UV. Reaction products of betulin with concentrated sulphuric acid were analysed by UV [46, 47].

MS. The fragmentation pattern of betulin [48] as well as its diacetate [49] was described. Further information about the mass spectra is found in a review article on pentacyclic triterpenes [50]. Molecular MS results in a simplified fragmentation pattern as expected [51].

Chemical reactivity.

Betulin has a high thermal stability [19]. On distillation at atmospheric pressure a Russia-leather [14] smell is observed. Kinetic investigations indicated a very low

reactivity of the hydroxy groups of betulin [52]. Oxidation, reduction and rearrangement reactions with acid catalysis were reviewed [4]. In addition, the homogeneous hydrogenation and deuteration [53] as well as an oxidation reaction by air [22] and a dehydrogenation by alkali fusion (NaOH-KOH) [54] are described. Using $\text{Hg}(\text{OAc})_2$ in HOAc-CHCl_3 as oxidant [55] an isomerisation product is observed [56]. The dehydration with POCl_3 leads to a Wagner-Meerwein rearrangement [57]. Catalytic glycosidation of betulin with several catalysts [58, 59] was studied and some betulin glycosides were synthesized [60, 61]. Tetraacetylglycopyranosylorthoesters of betulin have been prepared [62]. Other carbohydrate orthoesters underwent transesterification [63, 64]. The synthesis of betulin glucoside acetates via acyl halides [65] or orthoesters in the presence of HgBr_2 was studied [66, 67].

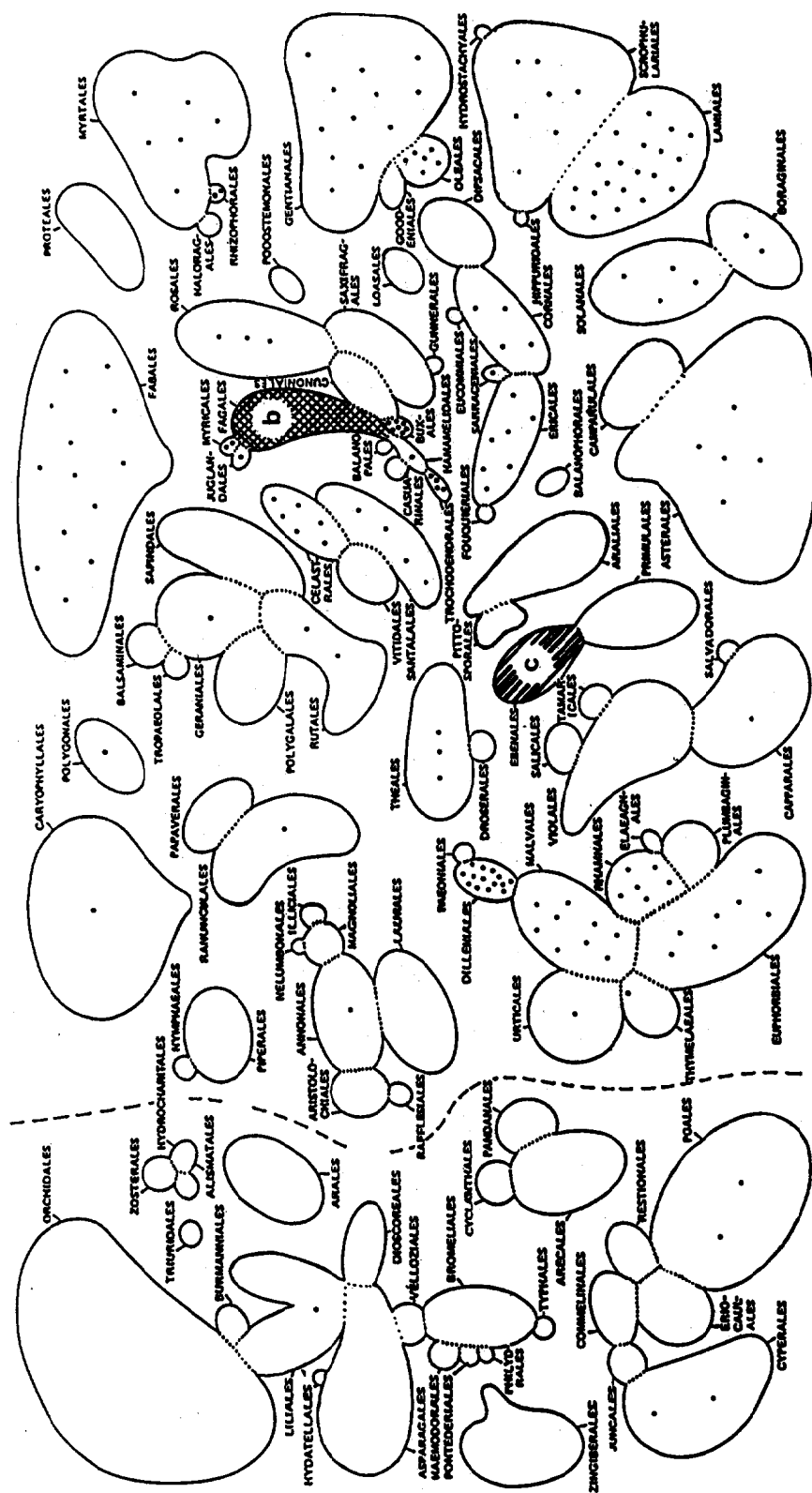


Fig. 1. Reported occurrence of betulin in angiosperms. (a) the number of dots reflects the number of betulin containing species. (b) the occurrence of betulin both quantitatively and qualitatively is concentrated within this order. (c) many investigations are reported for *Diospyros* and betulin was identified in 57 species. (Pedigree adopted from ref. [328].)

The absolute configuration of betulin was assigned by application of the Horeau-method and diastereoisomeric reaction products detected by GC [68, 69]. Betulin is presumed to have isomerized to allobetulin and then oxidized to hydroxyallobetulin in geological samples [70]. The biogenetic pathway to betulin has been shown to proceed via geraniol, farneosol and geranyl-geraniol to squalene [71, 72] which is further converted via lupeol [29] to betulin.

Uses

It has been estimated [22], that a pulp mill with an annual production capacity of 200 000 ton birch kraft pulp produces bark waste permitting isolation of ca 2500 ton per annum of betulin of 95% purity. Betulin, present in extractives of birchwood and pulp, is assumed to be the main cause of harmful resin deposits on papermaking machines [23] and was the main crystalline compound of the unsaponifiable fraction from hardwood sulphate soap in a Russian pulp mill [73]. It can be isolated from this source and the crude soap of the sulphate process [74]. Addition of 1% betulin solution to groundwood pulp handsheets increased lightfastness and decreased yellowing [75]; thus betulin can be used as light stabilizer for cellulose and wood pulp [76].

Acid betulin diphthalate [77] has been prepared and polyesters of betulin with linseed fatty acids yield quality protective coatings [78]. Furthermore betulin was used in the manufacture of resins, lacquers, emulsifiers, polyurethanes and in cosmetic products [4]. Polymers with rosin have good water resistance [79]. Polyurethanes with Desmodur E22 gave only oligomers [15]. Unsaturated polyesters had improved water and chemical resistance [52]. Polyacrylates of betulin were obtained in quantitative yield [80]. Betulin has inhibitory effects to the curing of a mixture of unsaturated resins [81].

Chemotaxonomy and distribution in the plant kingdom

The variation of the betulin content ranging from 5 to 22% in the leaves of four species of white barked birch is taxonomically useful [82]. Betulin containing plants are systematically widespread within the angiosperms and found predominantly among trees and bushes. An accumulation, both quantitatively and qualitatively, seems to be within the order Fagales; Reports on the occurrence of betulin are mainly dealing with the orders Buxales, Dilleniales, Ebenales and Lamiales as shown in the pedigree adopted from ref. [328] (Fig. 1). The distribution of betulin within families is depicted in Fig. 2.

The only occurrence of betulin in gymnosperms was reported for the American *Abies balsamea* [83]. Betulin was also identified in peat [16–18, 84–87], lignite [86] geochemical samples (e.g. kaolin [88] and rock wax [89, 90] as well as in fungi [91–95], kraft black liquors [96, 97], pulp [21, 23, 44, 74–76, 98, 99] and tall oil [100].

The identification of betulin in archaeological specimens gave evidence that tar produced from birch bark was used in prehistoric times as a multifunctional material, e.g. as glue and for paints coating the surface of clay vessels [13, 19, 101, 102]. Among the derivatives of betulin found in nature are betulin-3-cafate [28, 103], betulin diacetate and the 3 α -isomer *epi*-betulin [104] as well as some betulin glycosides [105, 106].

Pharmacology, toxicology, drugs

Birch bark (cortex *Betulae*) containing 10–14% betulin has been used as an antiseptic [107] and in folk medicine against skin diseases, malaria, dropsy and gout [108]. Similarly, birch tar (pix *Betulae*, oleum rusci) was used to treat rheumatism and gout [108] as well as a nematocide antiseptic [107] and against colic and mange in veterinary medicine. Birch bark oil is used as an antirheumatic and disinfectant [109]. Betulin showed inhibitory effects on Epstein-Barr virus activation [110] and bacteriostatic activities against *Escherichia coli*, *Salmonella typhi*, *Shigella flexneri* and *Staphylococcus aureus* [111]. It was tested against Walker 256 carcinosarcoma and MCF-7 human mammary adenocarcinoma *in vitro*, and leukemia P 388 *in vivo* and showed activity comparable to inotodiol [92]. Betulin has no effect on the growth of *Phytophthora cactorum* [112]. Betulin glycosides have no cytostatic effect in developing sea urchin embryos [105]. Betulin showed feeding deterrent activity [113]. Tumour inhibition with activity against human epidermoid carcinoma of the nasopharynx *in vitro* [114] and Walker 256 (SWA16) tumour system [115] but not against L-1210 lymphoid leukemia in mice or Walker carcinosarcoma 256 in rats [114] was reported. For betulin and betulin-containing herbs antiseptic [4, 116], antiinflammatory [117] and insecticidal activities have been mentioned.

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